Prepare test samples.

A protein standard is included in the kit. Prepare a protein standard of the target protein.

Add TMB Stop Solution

Duplicate assay wells are recommended for both standard and samples.

Subject the plate to analysis.

REFERENCES


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ELISA PROTOCOL

Preparation of Test Samples
1. Process Test Samples in the following manner:
   - **Cell culture supernate, tissue lysate or body fluids:** Remove particulates by centrifugation, analyze immediately or aliquot and store at -20°C.
   - **Serum:** Allow the serum to clot in a serum separator tube (about 30 min) at room temperature. Centrifuge at approximately 2000 X g for 20 min. Analyze the serum immediately or aliquot and store frozen at -20°C.
   - **Plasma:** Collect plasma using heparin as an anticoagulant. Centrifuge for 20 min at 2000 X g within 30 min of collection. Analyze immediately or aliquot and store frozen at -20°C.
2. Estimate the concentration of the target protein in the sample and select a proper dilution factor such that the diluted target protein concentration falls within the 156-10000 pg/ml standard curve range. Depending on the sample, several trial dilutions may be necessary. Dilute the sample using the provided diluent buffer, mixing well. Suggested working dilutions of standards are as follows:

<table>
<thead>
<tr>
<th>Target Protein Concentration Range</th>
<th>Sample Working Dilution</th>
<th>Sample Vol</th>
<th>Diluent Buffer Vol.</th>
</tr>
</thead>
<tbody>
<tr>
<td>100-1000 ng/ml</td>
<td>1:100</td>
<td>1 μl</td>
<td>99 μl</td>
</tr>
<tr>
<td>10-100 ng/ml</td>
<td>1:10</td>
<td>10 μl</td>
<td>90 μl</td>
</tr>
<tr>
<td>10-100 ng/ml</td>
<td>1:1</td>
<td>1 μl</td>
<td>99 μl</td>
</tr>
<tr>
<td>156-10000 pg/ml</td>
<td>1:2</td>
<td>50 μl</td>
<td>50 μl</td>
</tr>
<tr>
<td>≤156pg/ml</td>
<td>n/a</td>
<td>100μl</td>
<td>n/a</td>
</tr>
</tbody>
</table>
3. If samples will be assayed within 24 hours, store at 2-8°C. For long-term storage, aliquot and freeze samples at -20°C. Avoid repeated freeze-thaw cycles.

Preparation of Standard Solutions (156-10000 pg/ml)
4. Reconstitute the LypoPhorized Recombinant Protein to make a 10,000 pg/ml MMP-2 solution. Add 1 ml Sample Diluent Buffer to a tube of lyophiized protein, keep the tube at room temperature for 10 min. Mix thoroughly.
5. Label 6 eppendorf tubes with the MMP-2 protein concentrations to be prepared by serial dilution: 5000pg/ml, 2500pg/ml, 1250pg/ml, 625pg/ml, 313pg/ml, 156pg/ml.
6. Aliquot 0.3 ml of the Sample Diluent Buffer to the labeled tubes.
7. Serially dilute the protein standards into their respectively labeled tubes. Transfer 0.3 ml from the 10000pg/ml MMP-2 Solution to the 5000pg/ml eppendorf tube and mix thoroughly. Transfer 0.3 ml of the 5000 pg/ml solution to the 2500 pg/ml tube and mix thoroughly, Transfer 0.3 ml of the 2500 pg/ml solution to the 1250 pg/ml tube and mix, and so on to make the 625, 313 and 156 pg/ml solutions.
8. Store at 4°C until use.

Loading the 96-well Plate
9. Aliquot 0.1 ml of the sample diluent buffer into a control well to serve as the Blank. This will yield the O.D.450(Blank) reading.
10. Aliquot 0.1 ml of the standard solutions of the Preparation of Standard Solutions (156-10000pg/ml) into empty wells of the precoated 96-well plate. Duplicate measurements of standards are recommended.
11. Aliquot 0.1 ml of each properly diluted test sample to empty wells prepared in Step 2. Duplicate measurements of each test sample are recommended.
12. Cover the 96-well plate and incubate at 37°C for 90 min.
13. During the **Step 12** incubation period, prepare a stock of Biotinylated 1:100 Antibody Working Solution. Count the number of reactions and multiply by 0.1 ml/well for the Working Solution total volume (preparation of 1-2 reactions in excess of the number of wells is recommended). Dilute the Biotinylated Antibody to 1:100 in Antibody Diluent Buffer and mix thoroughly. Use the working solution within 2 hours.
14. Upon completion of the 90 min incubation of **Step 12**, remove the cover of the 96 well plate and discard plate well contents. Blot the plate onto paper towels or other absorbent material. DO NOT let the wells completely dry at any time.
15. Add 0.1 ml of the Biotinylated 1:100 Antibody Working Solution (prepared in **Step 13**) to each well and incubate the plate at 37°C for 60 min.
16. During the incubation period of **Step 15**, prepare a stock of ABC Working Solution. Count the number of reactions and multiply by 0.1 ml/well for the Working Solution total volume (preparation of 1-2 reactions in excess of the number of wells is recommended). Dilute the ABC Stock Solution to 1:100 in ABC Diluent Buffer and mix thoroughly. Pre-warm the ABC working solution at 37°C for 30 min before use. Use the working solution within 1 hour.
17. Upon completion of the 60 min incubation of **Step 15**, wash the plate 3 times with 0.3 ml TBS or PBS. For each wash, leave washing buffer in the wells for 1-2 min. Discard the washing buffer and blot the plate onto paper towels or other absorbent material.
18. Add 0.1 ml of prepared ABC Working Solution (prepared in **Step 16**) to each well and incubate the plate at 37°C for 30 min.
19. During the incubation period of **Step 18**, pre-warm TMB Color Developing Agent at 37°C for 30 min before use.
20. Upon completion of the 30 min incubation of **Step 18**, wash the plate 5 times with 0.3 ml TBS or PBS. For each wash, leave the washing buffer in the wells for 1-2 min. Discard the washing buffer and blot the plate onto paper towels or other absorbent material.
21. Add 90 μl of the pre-warmed TMB Color Developing Agent into each well and incubate at 37°C for 15-20 min (shades of blue can be seen in the wells with the four most concentrated Protein Standard Solutions; the other control wells should show no obvious color).
22. Add 0.1 ml of the TMB Stop Solution to each well. The acidic stop solution will change the mixture color to yellow. The yellow intensity is proportional to the amount of target protein captured by the plate.
23. Read the O.D. absorbance at 450nm in a microplate reader within 30 min after adding the stop solution. These readings are the O.D.450(Reading).

Calculating Protein Concentration
- For all wells, determine O.D.450(Relative):
  O.D.450(Relative) = O.D.450(Reading) – O.D.450(Blank)
- Plot the standard curve: plot O.D.450(Relative) of each standard solution (Y) vs. the respective concentration of the standard solution (X). See Figure 1 for a typical standard curve.
- The MMP-2 concentration of the samples can be interpolated from the standard curve. Multiply the interpolated concentration by the dilution factor to obtain the target protein concentration in the sample.

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